

WESTERN-BLOTTING PROTOCOL

**Electrophoresis and transfer** 

Separate proteins by gel electrophoresis according to manufacturer's specifications.

Run standard SDS-Polyacrylamide gel and electrophoreses in a 0.5-1.5 mm thick gel.

Transfer proteins from the gel to a nitrocellulose or other membrane such as PVDF by electro-blotting

according to the manufacturer's protocols.

**Blocking** 

Block remaining hydrophobic binding sites on the membrane by incubating membrane in a blocking

solution (5% non-fat dried milk, 0.05% Tween 20 in phosphate buffer solution for 30-60 minutes at

room temperature or overnight at 4°C.

**Primary Antibody Incubation** 

Dilute the primary antibody in the blocking buffer. Optimal antibody concentration should be

determined by titration (a concentration of 1-2.0  $\mu g/ml$  is generally acceptable). Incubate for 1 hour at

room temperature or overnight at 4°C with gentle agitation.

Wash membrane three times for 5 minutes each in washing buffer (phosphate buffer saline plus

0.05% Tween 20).

Secondary antibody incubation

Incubate the membrane with appropriate conjugated secondary antibody: alkaline phosphatase or

horseradish peroxydase conjugated antibody. Incubate the membrane with conjugated secondary

antibody diluted in blocking buffer for 30-60 minutes at room temperature with gentle agitation.

Wash membrane three times for 5 minutes each in washing buffer (phosphate buffered saline plus

0.05% Tween 20).

**Visualisation** 

Incubate the membrane with appropriate substrate solution and incubate for time recommended by

manufacturer to visualise protein bands.

**Solutions** 

**Blocking solution** 

5% (w/v) non-fat dried milk, 0.05% (v/v) Tween 20 in phosphate buffer solution

Washing buffer

Phosphate buffered saline plus 0.05% (v/v) Tween 20