Chick Embryo Extract, Ultrafiltrate, Liquid (CEE)

Catalog number

C3999

Supplier

United States Biological

Chick Embryo Extract, Ultrafiltrate is used as a supplement in some growth media formulations. It is prepared by blending 11-14 day old chick embryos in a balanced salt solution (3X embryo volume). The solution undergoes centrifugation to remove larger particles and debris. The supernatant is subjected to an ultrafiltration step with a 10kD MW cutoff to remove protein from the solution, producing a clear, amber liquid. This liquid is sterile-filtered.

Animals are sourced from a specific USDA-Registered facility using a veterinary-inspected flock. Final processing at an ISO9000 facility. Chick Embryo Extract, Ultrafiltrate is protein free. It does not support the growth of viruses.

Appearance

Pale-yellow, clear, solution

Total Protein (Biuret)

~0.0g/dL

Sterility (per 9CFR 113.26)

Negative

Mycoplasma (per 9CFR 113.26)

Negative

Osmolality

As Reported

Microbial Testing

Pullorum typhoid, Galisepticum, M. synoviae, M. meleagridis): Negative

Recommended Dilutions

1ml of extract/100ml of media

Storage and Stability

Chick Embryo Extract, Ultrafiltrate is protein-free; thus, it will not break down. May be stored at RT. Stable for 12 months after receipt. Storage at 4°C extends the stability for an additional 3-6 months. Some precipitate has been observed when frozen.

Important Note

This product as supplied is intended for research use only, not for use in human, therapeutic or diagnostic applications without the expressed written authorization of United States Biological.

Toxicity and Hazards

All products should be handled by qualified personnel only, trained in laboratory procedures.

Formulation

Pale-yellow, clear, solution

Grade

Molecular Biology Grade

Storage

4°C

Antigen Modification

11-14 day old chick embryos

Reference

US Biological Application References: 1. Griffin, M.A., et al., J. Cell Science 117: 5855-5863 (2004). 2. Clause, K.C., et al., Tissue Engineering Part C 16: 375-385 (2010). 3. Montemurro, T., et al., J. Cell Mol. Med. 15(4): 796–808 (2011). 4. Jiang, X., et al., Stem Cells Dev. 18: 1059-1070 (2009). 5. Lokireddy, S., et al., Am. J. Physiol. Cell Physiol. (2011) http://ajpcell.physiology.org/content/early/2011/09/01/ajpcell.00114.2011. 6. de la Garza-Rodea, A.S. et al., (2013) FASEB J. doi: 10.1096/fj.13-233155. 7. Park S., et al. 2014. J Exp Zool Mol Dev Evol. 322:156-165. 8. O'Connell G.C. and Pistilli E.E. Biochem Biophys Res Commun. 2015. 458:614-619. 9. Escobar H, et al. Molecular Therapy—Nucleic Acids (2016) 5, e277; doi:10.1038/mtna.2015.52. 10. Masuda S, et al. 2018. Acta Physiol (Oxf). 222(3). doi: 10.1111/apha.12975. Epub 2017 Oct 19. 11. Ahrens HE, et al. 2018. Skelet Muscle. 8(1):20. doi: 10.1186/s13395-018-0166-x.