

NZY Bis-Tris Precast Gel

(4-12%, 12 wells)

Catalogue number	MB46501
Presentation	10 gels
Gel percentage	4-12%
Well format	12 wells
Capacity	25 µL/well
Cassete size	Mini (10 x 8.3 cm)
Buffer system	MOPS and MES

Description

NZY Bis-Tris Precast Gel is a high-performance and user-friendly precast polyacrylamide gel designed for electrophoresis in the Bis-Tris buffer system (MOPS or MES). With its optimized formula, the NZY Bis-Tris Precast Gel exhibits enhanced resolution, accurate results, and an extended shelf-life compared to conventional Laemmli Tris-HCl gels.

The NZY Bis-Tris Precast Gels are available in a 4 to 12% gradient concentration, offering excellent separation capabilities. Precast gels come in a convenient 12-well format, making them ideal for various electrophoresis applications. These gels are offered in a Mini cassette size (10 x 8.3 cm) and are compatible with most popular protein electrophoresis systems such as Bio-Rad®.

Choose NZY Bis-Tris Precast Gels for superior performance, ease of use, and reliable results in your electrophoresis experiments.

Key Features

- Enhanced band sharpness
- Great resolution of small proteins
- Compatible with most popular protein electrophoresis systems
- Easy sample loading: numbered and framed wells
- Labelled warning sign and green tape as reminder
- Stable for shipping at room temperature

Standard Procedures

Gel Running recommendations

1. Before starting, remove both comb and tape.
2. Use fresh 1X running buffer for the inner cathode chamber.
3. Do not use Tris-Glycine running buffer for NZY Bis-Tris Precast Gels.
4. Prior to loading samples, rinse the wells.

Sample preparation for SDS-PAGE

1. Mix protein sample with 2X sample buffer.
2. Heat diluted samples at 95°C for 5 min or at 70°C for 10 min.
3. Cool the diluted samples to 4°C and spin down the water condensed on the tube surface. In case there is a high viscosity portion at the bottom of the tube, transfer the supernatant to a new tube.

Prepare Precast gel for sample loading

1. Open the blister tray of NZY Bis-Tris Precast Gel.
2. Briefly rinse the gel cassette with distilled H₂O.
3. Remove tape and comb; avoid squeezing the gel.
4. Adapt NZY Bis-Tris Precast Gel to the electrophoresis system. NZYtech recommends using BioRad Mini-PROTEAN® Core Electrophoresis System.
5. Use a pipette to gently wash the wells with running buffer to remove residual storage buffer.
6. Fill the wells with running buffer prior to sample loading.
7. Load samples and protein marker into numbered wells.
8. Fill both inner and outer chambers with running buffer to the highest level. Ensure gel wells are completely covered.

Gel running power configuration

VOLTAGE * ¹	130 V	180 V	230 V * ²
RUNNING TIME * ³	45-60 min	25-40 min	15-30 min
EXPECTED CURRENT			
Initial (per gel)	60-70 mA	100-110 mA	130-140 mA
Final (per gel)	20-25 mA	40-50 mA	60-70 mA
EXPECTED TEMPERATURE	25-30°C	25-35°C	35-45°C

*¹ It is recommended to set voltage higher than 100 V.

*² For higher voltage conditions, use fresh running buffer for inner and outer chambers.

*³ Running time varies depending on running buffer, temperature, and power supply.

If needed, optimize both voltage and running time.

Remove NZY Bis-Tris Precast Gel from Cassette

1. Open cassette immediately after electrophoresis. Avoid gel drying.
2. Insert the cassette opener into corners of cassette.
3. Gradually lever the opener to detach the two plates.
4. Gently pull two plates apart from the top of cassette.
5. Carefully detach the gel either from the bottom or the top side of the cassette. Avoid diagonally peeling the gel from the corner. Use water to help gel detachment if needed.
6. Gently remove the gel for further staining or Western blotting.

Gel Staining

Proteins separated using NZY Bis-Tris Precast Gel can be further stained with most popular staining reagents, such as BlueSafe (MB15201) or Coomassie dye.

Transferring Protein from NZY Bis-Tris Precast Gel to Blotting Membrane

1. After protein separation using NZY Bis-Tris Precast Gel, gently detach gel from cassette and then equilibrate it in transfer buffer.
2. Pre-soak blotting membrane and filter papers in transfer buffer.

Note: Activate PVDF membrane in methanol before soaking in transfer buffer. Prepare 6 filter papers for one gel/membrane sandwich.

3. Assemble transfer sandwich by orientating cathode, sponge, filter papers, gel, membrane, filter papers, sponge, and anode. The protein goes to the direction of cathode to anode.
4. Carefully move roller over the gel/membrane to remove air bubbles and excess buffer until complete contact is established.
5. Insert transfer cassette into transfer module. Notice that black side of cassette should be next to black side of module.
6. Fill transfer tank with pre-cooled transfer buffer to the highest water level.
7. Set constant voltage at 100 V. Transfer for 90 minutes at low temperature condition. Pre-stained protein marker should be visible on the membrane after transfer is completed. Transfer of proteins to the membrane can be checked using Ponceau S staining before blocking step.

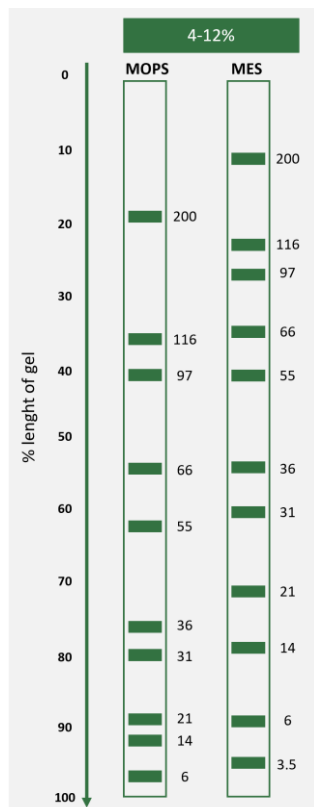
Shipping Conditions

This product must be shipped at room temperature.

Storage Conditions

Store NZY Bis-Tris Precast Gels at 2-8°C. Please do not freeze NZY Bis-Tris Precast Gels. During storage, it is essential to keep the NZY Bis-Tris Precast Gels flat to maintain their integrity and optimal performance. Keep the gels away from the cold air vent. Prior to electrophoresis, ensure to remove the tape and comb from the gels.

Migration pattern



Supplemental information

Adapting NZY Bis-Tris Precast Gel to BioRad Mini-PROTEAN® Core

1. After removing comb and tape, place the NZY Bis-Tris Precast Gel with notched plate facing toward inner chamber.
2. Align the notched plate to ensure the edge sits just below the notch at the top of green gasket.
3. Gently press gel cassette toward green gasket and then lock gel cassette with two green arms. Avoid squeezing the cassette and gel.
4. Fill inner chamber with running buffer to check tightness of seal. If necessary, reassemble and check the seal again.
5. Fill inner chamber with running buffer to ensure gel wells are completely covered.
6. Fill outer chamber with running buffer to the highest level.

Note: for other electrophoresis system, please follow the manufacturer's instructions

Buffer recipes

2X sample buffer with reducing agent

62.5 mM Tris-HCl pH 6.8, 2% SDS, 25% (v/v) glycerol, 0.01% bromophenol blue, 5% β -mercaptoethanol or 100 mM DTT (added fresh)

10X MOPS running buffer

60.6 g Tris base, 104.6 g MOPS, 10.0 g SDS, 3.0 g EDTA.

Bring up the volume to 1 L with ddH₂O.

10X MES running buffer

60.6 g Tris base, 97.6 g MES, 10.0 g SDS, 3.0 g EDTA.

Bring up the volume to 1 L with ddH₂O.

1X running buffer

Dilute 100 ml 10X running buffer with 900 ml ddH₂O.

10X transfer buffer

30.0 g Tris base, 144.0 g Glycine. Bring up the volume to 1 L with ddH₂O.

1X transfer buffer

Dilute 100 ml 10X transfer buffer with 200 ml methanol and 700 ml ddH₂O.

Note: Cool 1X transfer buffer to 4°C before using. Add SDS to 0.1% to promote transfer of high molecular weight proteins.

For life science research only. Not for use in diagnostic procedures.

NZYtech Lda. Estrada do Paço do Lumiar, Campus do Lumiar - Edifício E, R/C, 1649-038 Lisboa, Portugal Tel.:+351.213643514 Fax:
+351.217151168 www.nzytech.com

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Well format	12 wells
Capacity	25 µL/well
Cassette size	Mini gel (10 x 8.3 cm)
Gel dimensions	8.1 x 7.4 x 0.1 cm (width x length x thickness)
Buffer system	MOPS and MES

Description

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5. Use a pipette to gently wash the wells with running buffer to remove residual storage buffer.
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7. Load samples and protein marker into numbered wells.
8. Fill both inner and outer chambers with running buffer to the highest level. Ensure gel wells are completely covered.
9. Configure the electrophoresis system based on the table below. If needed, optimize both voltage and running time.

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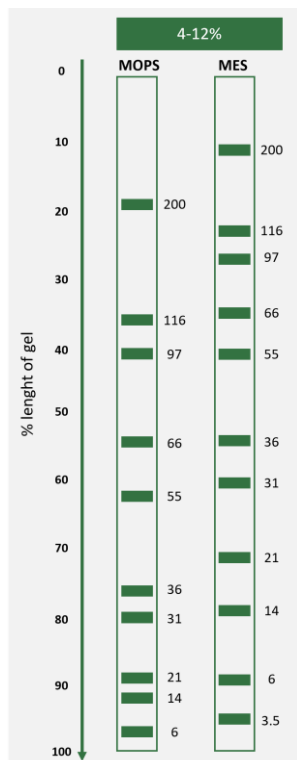
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