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**Configuration-specific Monoclonal Antibody Based**

**G $\alpha$ <sub>13</sub> Activation Assay Kit**

**(30 Assays)**

**Cat. # 80401**

**FOR RESEARCH USE ONLY. NOT FOR DIAGNOSTIC APPLICATIONS**

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# Gα<sub>13</sub> Activation Assay Kit Protocol

## Cat. # 80401

## Introduction

### A. Background

A structurally diverse repertoire of ligands, from photons to large peptides, activates GPCRs to elicit their physiological functions. Ligand-bound GPCRs, in turn, function as guanine nucleotide exchange factors catalyzing the exchange of GDP bound on the Gα subunit with GTP in the presence of Gβγ, causing the dissociation of the Gα subunit from the Gβγ dimer to form two functional units (Gα and Gβγ). Both Gα and Gβγ subunits signal to various cellular signaling pathways. Based on the sequence and functional homologies, G proteins are grouped into four families: G<sub>s</sub>, G<sub>i</sub>, G<sub>q</sub>, and G<sub>12</sub>. As increasing numbers of effectors and interacting proteins for these G proteins have been identified, the physiological processes in which G proteins participate are multiplying.

Among the four subfamilies of G proteins, the function of G<sub>12/13</sub> subfamily is less well understood. In this family, there are two members, G<sub>12</sub> and G<sub>13</sub>, that are expressed ubiquitously. Gα<sub>12</sub> knockout mice appeared normal. Gα<sub>13</sub> knockout mice displayed embryonic lethality (~E9.5). The Gα<sub>13</sub> -/- mouse embryos had defective vascular systems. G<sub>13</sub> is also essential for receptor tyrosine kinase-induced migration of fibroblast and endothelial cells.

### B. Assay Principle

NewEast Biosciences Gα<sub>13</sub> Activation Assay Kit uses configuration-specific anti-Gα<sub>13</sub>-GTP Mouse monoclonal antibody to measure Gα<sub>13</sub>-GTP levels either from cell extracts or from in vitro GTPγS loading Gα<sub>13</sub> activation assays. Anti-Gα<sub>13</sub>-GTP mouse monoclonal antibody is first incubated with cell lysates containing Gα<sub>13</sub>-GTP. Next, the GTP-bound Gα<sub>13</sub> is pulled down by protein A/G agarose. Finally, the precipitated Gα<sub>13</sub>-GTP is detected through immunoblot analysis using anti-Gα<sub>13</sub> mouse monoclonal antibody.

### C. Kit Contents

This kit contains enough reagents for approximately 30-35 pull-down assays.

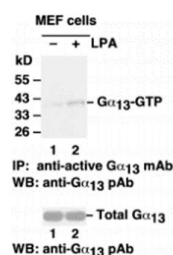
Reagent	Cat. #	Quantity	Storage
Anti-Gα <sub>13</sub> -GTP Mouse Monoclonal Antibody	26902	30 μL	-20°C
Protein A/G Agarose	30301	600 μL	4°C
5X Assay/Lysis Buffer	30303	30 mL	4°C
Anti-Gα <sub>13</sub> Mouse Monoclonal Antibody	21005	50 μL	-20°C
100X GTPγS	30302	50 μL	-20°C
100X GDP	30304	50 μL	-20°C
HRP-Goat Anti-Rabbit IgG	29002	50 μL	-20°C

### D. Materials Needed but Not Supplied

1. Stimulated and non-stimulated cell lysates
2. Protease inhibitors
3. 4°C tube rocker or shaker
4. 0.5 M EDTA at pH 8.0
5. 1.0 M MgCl<sub>2</sub>
6. 2X reducing SDS-PAGE sample buffer
7. Electrophoresis and immunoblotting systems
8. Immunoblotting wash buffer such as TBST (10 mM Tris-HCl, pH 7.4, 0.15 M NaCl, 0.05% Tween-20)
9. Immunoblotting blocking buffer (TBST containing 5% Non-fat Dry Milk or 3% BSA)
10. ECL Detection Reagents

### E. Example Results

The following figure demonstrates example results seen with the Gα<sub>13</sub> Activation Assay Kit. For reference only.



**Gα<sub>13</sub> Activation Assay.** MEF cells were treated with (lane 2) or without (lane 1) LPA. Cell lysates were incubated with an anti-Gα<sub>13</sub>-GTP monoclonal antibody (Cat. # 26902) (top panel). The precipitated active Gα<sub>13</sub> was immunoblotted with an anti-Gα<sub>13</sub> rabbit polyclonal antibody (Cat. # 21005). The bottom panel shows the Western blot with anti-Gα<sub>13</sub> of the cell lysates used (5% of that used in the top panel).

## Assay Procedure

### A. Reagent Preparation

**1X Assay/Lysis Buffer:** Shake the 5X Stock Buffer (Cat. # 30301) briefly and dilute with 4 times deionized water to make 1X buffer. Just prior to usage, add protease inhibitors such as 1 mM PMSF, 10 μg/mL leupeptin, and 10 μg/mL aprotinin.

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### B. Sample Preparation

#### Adherent Cells

1. Culture cells (one 10-cm plate,  $\sim 10^7$  cells) to approximately 80-90% confluence. Stimulate the cells with activator or inhibitor as desired.
2. Aspirate the culture media and wash twice with ice-cold PBS.
3. Completely remove the final PBS wash and add ice-cold 1X Assay/Lysis Buffer (See Reagent Preparation) to the cells (0.5–1 mL per 10 cm tissue culture plate).
4. Place the culture plates on ice for 10-20 minutes.
5. Detach the cells from the plates by scraping with a cell scraper.
6. Transfer the lysates to appropriate size tubes and place on ice.
7. If nuclear lysis occurs, the cell lysates may become viscous and difficult to pipette. If this occurs, lysates can be passed through a 27 $\frac{1}{2}$ -gauge syringe needle 3-4 times to shear the genomic DNA.
8. Clear the lysates by centrifuging at 12,000 x g and 4°C for 10 minutes.
9. Collect the supernatant and store the sample ( $\sim 1$ –2 mg of total protein) on ice for immediate use, or snap freeze and store at -70°C for future use.

#### Suspension Cells

1. Culture cells and stimulate with activator or inhibitor as desired.
2. Perform a cell count and then pellet the cells through centrifugation.
3. Aspirate the culture media and wash twice with ice-cold PBS.
4. Completely remove the final PBS wash and add ice-cold 1X Assay/Lysis Buffer (See Reagent Preparation) to the cell pellet (0.5-1 mL per  $10^7$  cells).
5. Lyse the cells by repeated pipetting.
6. Transfer the lysates to appropriate size tubes and place them on ice.
7. If nuclear lysis occurs, the cell lysates may become viscous and difficult to pipette. If this occurs, lysates can be passed through a 27 $\frac{1}{2}$ -gauge syringe needle 3-4 times to shear the genomic DNA.
8. Clear the lysates by centrifuging at 12,000 x g and 4°C for 10 minutes.
9. Collect the supernatant and store sample on ice for immediate use, or snap freeze and store at -70°C for future use.

### C. In vitro GTP $\gamma$ S/GDP Protein for Positive and Negative controls

*Note: In vivo stimulation of cells will activate approximately 10% of the available G $\alpha_{13}$ , whereas in vitro GTP $\gamma$ S protein loading will activate nearly 90% of G $\alpha_{13}$ .*

1. Aliquot 0.5 mL of cell extract (or 1  $\mu$ g of purified G $\alpha_{13}$  protein) into two microcentrifuge tubes.
2. To each tube, add 20  $\mu$ L of 0.5 M EDTA (final concentration of 20 mM).
3. Add 5  $\mu$ L of 100 X GTP $\gamma$ S (Cat. # 30302) to the first tube as a positive control.
4. Add 5  $\mu$ L of 100 X GDP (Cat. # 30304) to the second tube as a negative control.
5. Incubate both tubes at 30°C for 30 minutes with agitation.

6. Stop loading by placing the tubes on ice and adding 32.5  $\mu$ L of 1 M MgCl $_2$  (final concentration of 60 mM).

### D. Affinity Precipitation of Activated G Protein

1. Aliquot 0.5-1 mL of cell lysates (about 1 mg of total cellular protein) to a microcentrifuge tube.
2. Adjust the volume to 1 mL with 1X Assay/ Lysis Buffer (See Reagent Preparation).
3. Add 1  $\mu$ L anti-G $\alpha_{13}$ -GTP antibody (Cat. # 26902).
4. Prepare the protein A/G Agarose bead slurry (Cat. # 30301) by resuspending through vortexing or titrating.
5. Quickly add 20  $\mu$ L of resuspended bead slurry to above tube.
6. Incubate the tube at 4°C for 1 hour with gentle agitation.
7. Pellet the beads through centrifugation at 5,000 x g for 1 min.
8. Aspirate and discard the supernatant (making sure not to disturb or remove the bead pellet).
9. Wash the beads 3 times with 0.5 mL of 1X Assay/Lysis Buffer, centrifuging and aspirating each time.
10. After the third wash, pellet the beads through centrifugation and carefully remove all the supernatant.
11. Resuspend the bead pellet in 20  $\mu$ L of 2X reducing SDS-PAGE sample buffer.
12. Boil the sample for 5 minutes.
13. Centrifuge it at 5,000 x g for 10 seconds.

### E. Western Blot Analysis

1. Load 15  $\mu$ L/well of pull-down supernatant to a polyacrylamide gel (17%). It is recommended to include a pre-stained MW standard (as an indicator of a successful transfer in step 3 below).
2. Perform SDS-PAGE following the manufacturer's instructions.
3. Transfer the gel proteins to a PVDF or nitrocellulose membrane following the manufacturer's instructions.

*Note: Steps 4-11 are at room temperature with agitation*

4. Following electroblotting, immerse the PVDF membrane in 100% Methanol for 15 seconds, and then allow it to dry at room temperature for 5 minutes.

*Note: If Nitrocellulose is used instead of PVDF, step 4 should be skipped.*

5. Block the membrane with 5% non-fat dry milk or 3% BSA in TBST for 1 hr at room temperature with constant agitation.
6. Wash the blotted membrane three times with TBST, 5 minutes each time.
7. Incubate the membrane with anti-G $\alpha_{13}$  Mouse Monoclonal Antibody (Cat. # 21005), which is freshly diluted 1:50~500 (depending on the amount of G $\alpha_{13}$  proteins in your sample) in 5% non-fat dry milk or 3% BSA in TBST, for 1-2 hr at room temperature with constant agitation or at 4°C overnight.
8. Wash the blotted membrane three times with TBST, 5 minutes each time.
9. Incubate the membrane with a secondary antibody (Cat. # 29002), which is freshly diluted 1:1000 in 5% non-fat dry milk or 3% BSA in TBST, for 1 hr at room temperature with constant agitation.
10. Wash the blotted membrane three times with TBST, 5 minutes each time.
11. Use the detection method of your choice such as ECL.